Organization for Security and Co-operation in Europe

Date: 24 August 2023

CLARIFICATION NOTE #1

Reference: ITB/SEC/16/2023

Subject: Supply and delivery of laboratory reagents and consumables for veterinary labs in Ukraine

In accordance with issued **ITB/SEC/16/2023**, the OSCE would like to provide the following clarification to the ITB documents.

Please see **Annex A** on the next page where we have provided a specific nucleotide composition of required primers and probes. This complements technical requirements of **LOT 4**.

Please also note that according to this Clarification Note **the deadline for submission of bids** for opened ITB **is extended until 08 September 2023, 18:00 Vienna time (CET).**

Sincerely,

Dmytro Agapov,
Associate Procurement Officer,
OSCE Secretariat

ANNEX A

LOT 4 – SUPPLY AND DELIVERY OF OLIGONUCLEOTIDES AND PROBES

LIST OF REQUIRED PCR PRIMERS AND PROBES

No.	Name	Sequence	Comment
1.	Classical Swine Fever primers	CSF 100-F: 5'-ATG-CCC-AYA-GTA-GGA-CTA-GCA-3'	
2.	Classical Swine Fever primers	CSF 192-R: 5'-CTA-YTG-ACG-ACT-RTC-CTG-TAC-3'	
3.	Classical Swine Fever primers	CSF probe 5'-FAM-TGG-CGA-GCT-CCC-TGG-GTG-GTC-TAA-GT-TAMRA-3' (or BHQ1-3')	
4.	Rabies primers	RV F GATCCTGATGAYGTATGTTCCTA	
5.	Rabies primers	RV R RGATTCCGTAGCTRGTCCA	
6.	Rabies primers	RV probe FAM-CAGCAATGCAGTTYTTTGAGGGGAC-BHQ1	
7.	Leptospira primers	LipgrF2, 5'CGCTGAAATGGGAGTTCGTATGATTTCC3'	
8.	Leptospira primers	LipgrR2, 5'GGCATTGATTTTTCTTCYGGGGTWGCC3'	
9.	Leptospira primers	LipgrP1, 5'FAM AGGCGAAATCGGKGARCCAGGCGAYGG3'BHQ1	
10.	Salmonella primers	ttr-6 (forward), CTCACCAGGAGATTACAACATGG	
11.	Salmonella primers	ttr-4 (reverse), AGCTCAGACCAAAAGTGACCATC	
12.	Salmonella primers	Target probe (ttr-5), FAM-CACCGACGGCGAGACCGACTTT-Dark Quencher	
13.	Listeria primers	inlA_F, TCGCAAACAGATCTAGACCAAGTT	
14.	Listeria primers	inlA_R, GTTCAAGTATTCCAATCCATCGATAG	
15.	Listeria primers	inlA_P, FAM-CAACGCTTCAGGCGGATAGATTAGGGAT-TAMRA	
16.	Anthrax primers	dhp61 forward: 5'CGTAAGGACAATAAAAGCCGTTGT	
17.	Anthrax primers	dhp61 reverse: 5'CGATACAGACATTTATTGGGAACTACAC	
18.	Anthrax primers	dhp61 probe: 5'-6FAM-TGCAATCGATGAGCTAATGAACAATGACCCT-TMR	
19.	Anthrax primers	pagA forward: GTACAAGTGCTGGACCTACG	
20.	Anthrax primers	pagA reverse: CACTGTACGGATCAGAAGCC	
21.	Anthrax primers	pagA probe: FAM-ACCGTGACAATGATGGAATCCCTGA-BBQ	
22.	Anthrax primers	capC forward: CCTGCAGGTTTAGTTGTACCT	
23.	Anthrax primers	capC reverse: ACCTGTAATTAGCGTTGCCG	
24.	Anthrax primers	capC probe: FAM-AGCACTCGTTTTTAATCAGCCCGT-BBQ	
25.	Anthrax primers	gyrA forward: ATGTCAGACAATCAACAACAAGC	
26.	Anthrax primers	gyrA reverse: GCAATGAGTGTTATCGTATCTCG	
27.	Anthrax primers	gyrA probe: FAM-TATTAGCCATGAAATGCGTACCTC-BBQ	
28.	F. turarensis primers	fopAF: ATCTAGCAGGTCAAGCAACAGGT	

29.	F. turarensis primers	fopAR: GTCAACACTTGCTTGAACATTTCTAGATA	
30.	F. turarensis primers	fopAP: 6-FAM-CAAACTTAAGACCACCACCCACATCCCAA-BHQ-1	
31.	Newcastle disease virus	NDV_F, GGTGAGTCTATCCGGARGATACAAG	
32.	Newcastle disease virus	NDV_R, AGCTGTTGCAACCCCAAG	
33.	Newcastle disease virus	NDV_P, FAM-AAGCGTTTCTGTCTCCTTCCTCCA-TMR	
34.	AIV, M-gene	sep1, AGATGAGTCTTCTAACCGAGGTCG	
35.	AIV, M-gene	sep2, TGCAAAAACATCTTCAAGTCTCTG	
36.	AIV, M-gene	sep-probe, FAM-TCAGGCCCCCTCAAAGCCGA-TMR	
37.	AIV-H5 subtype	H5LH1, ACATATGACTACCCACARTATTCAG	
38.	AIV-H5 subtype	H5RH1, AGACCAGCTAYCATGATTGC	
39.	AIV-H5 subtype	H5PRO, FAM-TCWACAGTGGCGAGTTCCCTAGCA-TMR	
40.	AIV-H7 subtype	H7-F, CAACTGAAACRGTRGARCG	
41.	AIV-H7 subtype	H7-P,FAM-CCCAGGATYTGCTCAARAGGRAAAA-BHQ1	
42.	AIV-H7 subtype	H7-R1, CAGGAGYCCACATTGACC	
43.	AIV-H7 subtype	H7-R2, CAGWAGYCCACATTGACC	
44.	AIV-H7 subtype	H7-R3, TTCTAGGAATTGGTCACATTG	
45.	AIV-N1 subtype	N1 for, TAYAACTCAAGGTTTGAGTCTGTYGCTTG	
46.	AIV-N1 subtype	N1 rev, ATGTTRTTCCTCCAACTCTTGATRGTGTC	
47.	AIV-N1 subtype	N1-Probe, HEX-TCAGCRAGTGCYTGCCATGATGGCA-BHQ	
48.	AIV-N2 subtype	IAV-N2-1367F, AGTCTGGTGGACYTCAAAYAG	
49.	AIV-N2 subtype	IAV-N2-1488R, AATTGCGAAAGCTTATATAGVCAT	
50.	AIV-N2 subtype	N2-P, FAM-CCATCAGGCCATGAGCCT-MGB	
51.	AIV-N3 subtype	IAV-N3-1348F, AAYAGTATAGTTACTTTCTGYGG	
52.	AIV-N3 subtype	IAV-N3-1422R, CCAATGTTRGAACCATCHGG	
53.	AIV-N3 subtype	IAV-N3-1373P, FAM-TARACAATGAACCTGGATCGGGVAA-BHQ1	
54.	AIV-N7 subtype	IAV-N7-1305F, GTTGAATTAATWAGAGGAAGRCC	
55.	AIV-N7 subtype	IAV-N7-1430R, GATYTGTGCCCCATCRGGGA	
56.	AIV-N7 subtype	IAV-N7-1383P, FAM-AGCCCADTCYCAGTTGGGTCYGGTTC-BHQ1	
57.	AIV-N8 subtype	IAV-N8-1296F, TCCATGYTTTTGGGTTGARATGAT	
58.	AIV-N8 subtype	IAV-N8-1423R, GCTCCATCRTGCCAYGACCA	
59.	AIV-N8 subtype	IAV-N8-1354P, FAM-TCHAGYAGCTCCATTGTRATGTGTGGAGT-BHQ1	
60.	Francisella tularensis	Tul4_F, ATTACAATGGCAGGCTCCAGA	
61.	Francisella tularensis	Tul4_R, TGCCCAAGTTTTATCGTTCTTCT	
62.	Francisella tularensis	Tul4_P, FAM-TTCTAAGTGCCATGATACAAGCTTCCCAATTACTAAG-BHQ	

63.	Brucella spp	IS711_R, GGCCTACCGCTGCGAAT	
64.	Brucella spp	IS711_P, FAM-AAGCCAACACCCGGCCATTATGGT-BHQ	
65.	Bacillus anthracis	dhp61_F, 5‘CGTAAGGACAATAAAAGCCGTTGT	chromosome marker
66.	Bacillus anthracis	dhp61_R, CGATACAGACATTTATTGGGAACTACAC	chromosome marker
67.	Bacillus anthracis	dhp61_P, FAM-TGCAATCGATGAGCTAATGAACAATGACCCT-TMR	chromosome marker
68.	Bacillus anthracis	pagA_F, GTACAAGTGCTGGACCTACG	plasmid marker pXO1
69.	Bacillus anthracis	pagA_R, CACTGTACGGATCAGAAGCC	plasmid marker pXO1
70.	Bacillus anthracis	pagA_P, FAM-ACCGTGACAATGATGGAATCCCTGA-BBQ	plasmid marker pXO1
71.	Bacillus anthracis	capC_F, CCTGCAGGTTTAGTTGTACCT	plasmid marker pXO2
72.	Bacillus anthracis	capC_R, ACCTGTAATTAGCGTTGCCG	plasmid marker pXO2
73.	Bacillus anthracis	capC_P, FAM-AGCACTCGTTTTTAATCAGCCCGT-BBQ	plasmid marker pXO2
74.	Bacillus anthracis	GAPDH_F, CCACCCATGGCAAATTCC	internal control
75.	Bacillus anthracis	GAPDH_R, TCGCTCCTGGAAGATGGTG	internal control
76.	Bacillus anthracis	GAPDH_P, ROX-TGGCACCGTCAAGGCTGAGAACGT-BHQ2-3	internal control